

Large Animal Candidate Boot Camp October 20-23, 2022 University of Georgia, Veterinary Education Center | Athens, GA

Premier Sponsor

Boehringer Ingelheim

DAY 1 Thursday, October 20 th		
Time (Eastern Time)	Торіс	Presenter(s)
7:30 am	Shuttle to Veterinary Education Center (VEC)	
7:45-8:05 am	Registration	
8:05-8:10 am	Welcome & Introductions	Dr. Michelle Barton
8:10-9:00 am	 BAL and TTW Discuss indications for performing a TTW or a BAL Discuss proper technique for performing TTW and BAL 	Dr. Clare Ryan
9:10-10:00 am	 Clinical Neurologic Exam Understand how to perform a complete neurologic exam in horses and other large animals Interpret the results of the exam to perform accurate lesion localization and determine logical subsequent diagnostics steps 	Dr. Kelsey Hart
10:10-11:00 am	Cystoscopy and Urine Collection Perform urethral/bladder catheterization in both horse sexes Perform ureteral catheterization in mares Prepare an equine patient for and perform cystoscopy Recognize indications for performing these procedures	Dr. Hal Schott
11:10-12:00 pm	Goat Abdominal Ultrasonography	Dr. Brent Credille
12:00-1:00 pm	Lunch	









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DAY 1 (continued) Thursday, October 20 th		
Time (Eastern Time)	Торіс	Presenter(s)
1:00-3:00 pm	 Labs – 60 minutes each rotation Lab 1: Goat Abdominal Ultrasonography Identify the normal ultrasonographic abdominal anatomy of goats Identify common abdominal ultrasonographic abnormalities of goats through case examples Lab 2: Neurologic Exam Perform an advanced examination of the equine neurologic system Identify and localize neurologic abnormalities during a live "mystery case" examination Lab 3: BAL Know the materials needed for a BAL Perform a BAL on a standing sedated horse Lab 4: Urethral/Ureteral Catheterization (Mare) Obtain urine from the urinary bladder and ureters of a mare Perform cystoscopy and identify normal structures of the lower urinary tract of a mare Perform a urinary bladder mucosal biopsy (mare or gelding lab) 	Dr. Brent Credille, Dr. Kelsey Hart, Dr. Clare Ryan, Dr. Katie Mullen, Dr. Hal Schott, Dr. Wendy Vaala
3:00-3:30 pm	Break	
3:30-5:30 pm	Labs – 60 minutes each rotation Lab 1: Goat Abdominal Ultrasonography Lab 2: Neurologic Exam Lab 3: BAL Lab 4: Urethral/Ureteral Catheterization (Mare)	Dr. Brent Credille, Dr. Kelsey Hart, Dr. Clare Ryan, Dr. Katie Mullen, Dr. Hal Schott, Dr. Wendy Vaala
5:30-5:45 pm	Break	
5:45-6:30 pm	Tour of UGA Large Animal Veterinary Teaching Hospital	Dr. Michelle Barton
6:30-8:30 pm	Dinner – Clinical Case Discussions	
8:30 pm	Shuttle to Hotel	









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DAY 2 Friday, October 21⁵t		
Time (Eastern Time)	Торіс	Presenter(s)
7:45 am	Shuttle to Veterinary Education Center (VEC)	
8:00-8:50 am	 Gastroscopy Understand the application of the equine gastroscopic examination in assessment of the equine patient Describe the utility of the equine gastroscopic examination Illustrate the process of performing an equine gastroscopy in a thorough and efficient manner 	Dr. Michelle Coleman
9:00-9:50 am	 Echocardiography – The Basics List general indications for an echocardiogram in large animals Describe how to perform an echocardiogram on a large animal Describe common cardiac conditions and the echocardiographic findings that support or confirm the diagnosis 	Dr. Erin Beasley
9:50-10:10 am	Break	
10:10-12:10 pm	 Labs – 60 minutes each rotation Lab 5: Echocardiography Identify the normal ultrasonographic anatomy of the equine heart Obtain the standard right and left-sided echocardiographic views of the equine heart Lab 6: Cystoscopy (Gelding) Obtain urine from the urinary bladder and ureters of a gelding Perform cystoscopy and identify normal structures of the lower urinary tract of a gelding Perform a urinary bladder mucosal biopsy (mare or gelding lab) Lab 7: TTW Know the materials and alternative materials needed to perform a TTW Perform a TTW on a standing sedated horse Lab 8: Neurologic Case Reviews Detect neurologic abnormalities in large animals Perform accurate neuroanatomic lesion localization Interpret neurologic exam findings in light of patient signalment, history, and lesion localization to determine a list of relevant differential diagnoses	Dr. Erin Beasley, Dr. Michelle Barton, Dr. Hal Schott, Dr. Wendy Vaala, Dr. Clare Ryan, Dr. Katie Mullen, Dr. Kelsey Hart





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DAY 2 (continued) Friday, October 21 st		
Time (Eastern Time)	Торіс	Presenter(s)
12:10-1:10 pm	Lunch	
1:10-2:00pm	 Drug Use in Companion Food and Fiber Animals Understand AMDUCA and how it impacts pharmaceutical use in food and fiber animals Understand the principles of extra-label drug use Understand pharmaceutical agents that have restricted uses in food-producing animals Demonstrate an understanding of the application of extra-label drug use principles to clinical cases using case examples 	Dr. Brent Credille
2:10-3:00 pm	 Thoracocentesis and Chest Tube Placement Recognize pleural effusion ultrasonographically Recognize the presence of pneumothorax How to perform an ultrasound guided thoracocentesis Know the indications for chest tube placement 	Dr. Michelle Barton
3:00-3:30 pm	Break	
3:30-5:30 pm	Labs – 60 minutes each rotation Lab 5: Echocardiography Lab 6: Cystoscopy (Gelding) Lab 7: TTW Lab 8: Neurologic Case Reviews	Dr. Erin Beasley, Dr. Michelle Barton, Dr. Hal Schott, Dr. Wendy Vaala, Dr. Clare Ryan, Dr. Katie Mullen, Dr. Kelsey Hart
5:30-6:00 pm	Break	
6:00-8:00 pm	Dinner – Clinical Case Discussions	
8:30 pm	Shuttle to Hotel	









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DAY 3 Saturday, October 22 nd		
Time (Eastern Time)	Торіс	Presenter(s)
7:45 am	Shuttle to Veterinary Education Center (VEC)	
8:00-8:50 am	 C1C2/LS CSF Collection Describe how to perform CSF collection in standing horses Discuss indications, contraindications, and possible complications of cerebrospinal fluid collection 	Dr. Kelsey Hart, Dr. Erin Beasley
9:00-9:50 am	 Skin Scrapes and Biopsy Know the materials needed for a skin scraping and skin biopsy Know the 5 different types of skin biopsies Prepare cytologic specimens for submission to a diagnostic laboratory 	Dr. Suzi White
9:50-10:10 am	Break	
10:10-12:10 pm	 Labs – 60 minutes each rotation Lab 9: LS CSF Collection (Equine) Know the materials needed for LS CSF collection in the horse Know the anatomical landmarks for performing a LS CSF collection Perform LS CSF collection on a standing sedated horse Lab 10: C1C2 CSF Collection Know the materials needed for C1C2 CSF collection in the horse Identify the C1C2 CSF collection site ultrasonographically Perform C1C2 CSF collection on a standing sedated horse Lab 11: Gastroscopy and Biopsy Identify normal anatomy of the equine stomach, antrum, pylorus and proximal duodenum Perform gastroscopy and duodenoscopy on a standing sedated horse Obtain gastric and/or duodenal biopsies on a standing sedated horse Obtain gastric and/or duodenal biopsies on a standing sedated horse Perform a skin scrape Perform a skin scrape Perform the various methods of obtaining skin biopsies in horses Lab 13: Thoraccentesis Know the materials needed for thoracocentesis in the horse Perform a thoracocentesis in a standing sedated horse 	Dr. Kelsey Hart, Dr. Katie Mullen, Dr. Erin Beasley, Dr. Clare Ryan, Dr. Michelle Coleman, Dr. Suzi White, Dr. Michelle Barton









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DAY 3 (continued) Saturday, October 22 nd		
Time (Eastern Time)	Торіс	Presenter(s)
12:10-1:10 pm	Lunch	
1:10-2:00pm	 Atrial Fibrillation and Case Selection for Conversion Identify cases that are candidates for cardioversion Describe the differences between pharmacologic and transvenous electrical cardioversion Understand the risks and limitations for athletic potential in horses with permanent atrial fibrillation and the recurrence rate for horses that have had successful cardioversion therapy 	Dr. Erin Beasley
2:10-3:00 pm	 Common Skin Abnormalities Recognize common equine skin abnormalities Be able to make a rule out list for common skin abnormalities 	Dr. Suzi White
3:00-3:30 pm	Break	
3:30-5:30 pm	Labs – 60 minutes each rotation Lab 9: LS CSF Collection (Equine) Lab 10: C1C2 CSF Collection Lab 11: Gastroscopy and Biopsy Lab 12: Skin Scrapes and Biopsy Lab 13: Thoracocentesis	Dr. Kelsey Hart, Dr. Katie Mullen, Dr. Erin Beasley, Dr. Clare Ryan, Dr. Michelle Coleman, Dr. Suzi White, Dr. Michelle Barton
5:30-6:00 pm	Break	
6:00-8:00 pm	Optional Offsite Dinner - TBD	









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DAY 4 Sunday, October 23 rd		
Time (Eastern Time)	Торіс	Presenter(s)
7:45 am	Shuttle to Veterinary Education Center (VEC)	
8:00-8:50 am	 AO CSF Collection and Myelograms Describe how to safely perform CSF collection from the AO space in anesthetized horse Describe how, and why, to perform cervical myelography in anesthetized horses Discuss indications, contraindications and possible complications of these procedures 	Dr. Kelsey Hart
8:50-9:15 am	Break	
9:15-10:15 am	 Labs – 60 minutes each rotation Lab 14: AO CSF Collection/Myelogram Demonstration (Equine) Know the materials needed for an AO tap and myelogram in the horse Identify the anatomical landmarks for doing an AO tap in the horse Know how to do a myelogram Lab 15: TVEC Equipment/Procedure Review and Case Discussion Know the equipment involved in and the principles of how to perform TVEC in a horse Confirm intracardiac electrode placement radiographically through case discussion Lab 16: LS CSF Collection (Caprine) Know the anatomical landmarks for performing a LS CSF collection in goats Perform LS CSF collection on a goat 	Dr. Kelsey Hart, Dr. Clare Ryan, Dr. Amanda Coleman, Dr. Brent Credille









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DAY 4 (continued) Sunday, October 23 rd		
Time (Eastern Time)	Торіс	Presenter(s)
10:15-10:40 am	Break	
10:40-11:40 am	Labs – 60 minutes each rotation Lab 14: AO CSF Collection/Myelogram Demonstration (Equine) Lab 15: TVEC Equipment/Procedure Review and Case Discussion Lab 16: LS CSF Collection (Caprine)	Dr. Kelsey Hart, Dr. Clare Ryan, Dr. Amanda Coleman, Dr. Brent Credille
11:40-12:15 pm	Break	
12:15-1:15 pm	Lunch and Course Wrap-Up	





