

Large Animal Candidate Boot Camp
September 25-28, 2025
Purdue University | West Lafayette, IN.

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DAY 1 Thursday, September 25		
Time (Eastern Time)	Topic	Presenter(s)
7:30 am	Shuttle to Brunner Equine Hospital (Purdue University)	
7:45-8:00 am	Registration	
8:00-8:30 am	Welcome & Introductions	Taylor
8:30-9:00 am	Tour	Taylor
9:00-9:40 am	Equine Gastric Ulcer Syndrome <ul style="list-style-type: none"> Understand the differences in pathophysiology between ESGD and EGGD Assign a score to various gastric lesions Identify risk factors for development of EGUS Develop treatment plan for ESGD and EGGD 	Taylor
9:40-10:20 am	Guttural Pouch Disease <ul style="list-style-type: none"> Identify important anatomical structures within the guttural pouches Create differential diagnosis list for guttural pouch empyema Discuss treatment strategies for <i>Streptococcus equi</i> spp. <i>equi</i> infection 	Trsan
10:20-10:40 am	Break	
10:40-11:30 am	Equine Muscle Diseases and Principles of Biopsy <ul style="list-style-type: none"> Identify clinical presentations and indications for muscle biopsy in horses Understand the significance of different processing techniques for muscle biopsy Identify the sites for different muscle biopsies and discuss relevance of correct biopsy site Understand the practical approach to muscle biopsies and prepare for lab-based skills 	Jamieson

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DAY 1 (continued) Thursday, September 25		
Time (Eastern Time)	Topic	Presenter(s)
11:30 am-12:15 pm	Equine Theriogenology for the LAIM Specialist <ul style="list-style-type: none"> Diagnose placentitis Assess fetal health Identify safe and proper medications for the pregnant mare Diagnose and treat peri-partum conditions 	Hayna
12:15-1:00 pm	Lunch	
1:00-5:30 pm (Break 2:30-3:00 pm)	Labs – 50 minutes per rotation Lab 1: Gastroscopy <ul style="list-style-type: none"> Pass gastroscope correctly Identify relevant anatomical structures include greater and lesser curvatures, cardia, and pylorus If applicable, score ESGD and/or describe EGGD lesions Lab 2: Guttural Pouch Endoscopy <ul style="list-style-type: none"> Identify relevant anatomical structures for proper endoscopy of GP Perform bilateral GP endoscopy Pass uterine pipette into GP opening using endoscopy guidance Lab 3: Muscle biopsy <ul style="list-style-type: none"> Review sites for muscle biopsies taken in horses Understand procedure prep required to safely perform muscle biopsy in horses Perform or assist with gluteal, SCDM and semimembranosus biopsy Perform post procedure care for the patient and sample handling Understand and discuss how to mitigate common pitfalls associated with muscle biopsy Lab 4: Reproductive Ultrasound <ul style="list-style-type: none"> Measure CTUP with transrectal and transabdominal ultrasound Identify fetal heartbeat with M-mode ultrasound in pregnant mare 	Taylor, Trsan, Jamieson, Hayna

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DAY 1 (continued) Thursday, September 25		
Time (Eastern Time)	Topic	Presenter(s)
5:30-7:30 pm	Dinner on-site: Career Paths in LAIM	<i>Faculty</i>
7:30 pm	Shuttle to hotel	

DAY 2 Friday, September 26		
Time (Eastern Time)	Topic	Presenter(s)
7:30 am	Shuttle to Brunner Equine Hospital (Purdue University)	
8:00-8:50 am	Ruminant Forestomach Disease <ul style="list-style-type: none"> • Review anatomy of the ruminant forestomach • Understand the pathophysiology and clinical signs of common disorders of the ruminant forestomach <ul style="list-style-type: none"> ○ Bloat ○ Vagal indigestion ○ Ruminal acidosis 	<i>Bornheim</i>
8:50-9:50 am	Farm Animal Antimicrobial use <ul style="list-style-type: none"> • Understand essential farm animal drug laws for clinical practice • Define ELDU and navigate ELDU decision algorithm • Utilize FARAD to develop meat and milk withdrawals for ELDU • Identify main farm animal antibiotic classes and those with restricted use 	<i>Heller</i>

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DAY 2 (continued) Friday, September 26		
Time (Eastern Time)	Topic	Presenter(s)
9:50-10:10 am	Break	
10:10-11:00 am	Blood Collection and Restraint (Physical & Chemical) in Pet Pigs <ul style="list-style-type: none"> Develop appreciation for pet pig care, husbandry, and common healthy problems Become proficient with venipuncture in pet pigs Become familiar with chemical restraint options in pet pigs Become proficient in the safe handling and restraint of pet pigs 	<i>Ragland</i>
11:00-12:00 pm	Small Ruminant Urinary Tract Disorders <ul style="list-style-type: none"> Review predisposing factors for the development of urinary tract obstructions Identify common uroliths found in ruminants Review the anatomy of the urinary tract and note common areas of urinary obstruction Review medical and surgical treatment options for obstructive urolithiasis 	<i>Bornheim</i>
12:00-1:00 pm	Lunch: Sanctuary Ethics	<i>Ragland</i>

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1:00-5:30 pm (Break 2:30-3:00 pm)	<p>Labs – 50 minutes per rotation</p> <p>Lab 5: Ruminant Ultrasound</p> <ul style="list-style-type: none"> • Develop competence in ruminant point of care/rapid ultrasound (FLASC US) • Identify the 7 sites of a FLASC US in a cow and the normal anatomy at each • Understand expected abnormalities in each location and interpret their clinical significance <p>Lab 6: Small Ruminant Urethral Process Removal</p> <ul style="list-style-type: none"> • Review sedation protocols for urethral process removal • Safely rump sheep for non-sedated restraint and evaluation of the male urinary and reproductive tract • Remove the urethral process from intact or castrated small ruminants <p>Lab 7: Rumen Fluid Analysis</p> <ul style="list-style-type: none"> • Collect rumen fluid from donor cow through cannula • Pass orogastric tube • Determine rumen pH, perform methylene blue reduction test, and assess protozoa diversity and motility <p>Lab 8: Pig Restraint and Blood Collection +/- ear catheter placement</p> <ul style="list-style-type: none"> • Safely handle and restrain pet pigs • Perform venipuncture in pet pigs • Catheterize ear vessels of pet pigs 	Jamieson, Bornheim, Heller, Ragland
5:45 pm	Shuttle to hotel	

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DAY 3 Saturday, September 27		
Time (Eastern Time)	Topic	Presenter(s)
7:30 am	Shuttle to Brunner Equine Hospital (Purdue University)	
8:00-8:50 am	Equine Endocrinology <ul style="list-style-type: none"> Understand steps involved in assessment of equine endocrine system Develop a diagnostic plan for cases at risk of equine endocrine disorders Create a dietary plan for horses with EMS and PPID Understand the pharmaceutical management of equine endocrine disorders 	<i>Bertin</i>
8:50-9:40 am	Equine Neurologic Exam & CSF Collection <ul style="list-style-type: none"> Understand the steps involved in an equine neurologic evaluation Localize lesions based on clinical presentation Describe the equine neurologic gait deficit grading scale Describe the C1/C2, LS, and AO CSF collection procedures 	<i>Taylor</i>
9:40-10:00 am	Break	
10:00-11:00 am	Fluid Therapy in Horses <ul style="list-style-type: none"> Understand how to calculate fluid deficit, maintenance, and ongoing losses Create fluid therapy plans based on clinical presentation focusing on specific clinical challenges <ul style="list-style-type: none"> Liver failure Myopathies Post transfusion electrolyte management Hypoproteinemia Perform sodium correction calculations Prepare and calculate needs for isotonic bicarbonate solution 	<i>Jamieson</i>

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DAY 3 (continued) Saturday, September 27		
Time (Eastern Time)	Topic	Presenter(s)
11:00 am - 12:00 pm	Equine Ophthalmology <ul style="list-style-type: none"> Interpret abnormalities of an ophthalmological exam and interpret diagnostic test results (fluorescein and Rose Bengal staining, corneal cytology, tonometry, ocular ultrasonography) Formulate an appropriate treatment, prognosis, prevention, and monitoring plan for common ophthalmic conditions (corneal ulcers, stromal abscesses, uveitis) Understand indications for sub-palpebral lavage systems and intravitreal injections 	Townsend
12:00-1:00 pm	Lunch	
1:00-5:30 pm (Break 2:30-3:00 pm)	Labs – 50 minutes per rotation Lab 9: Equine Endocrine Case Discussions (BREQ, Fessler) <ul style="list-style-type: none"> Describe testing protocols for common equine endocrine disorders Interpret results of hormonal testing Assess the response to management using clinical and clinicopathological data Lab 10: Equine C1/C2 or L/S CSF Collection <ul style="list-style-type: none"> List the materials needed for each CSF collection Identify the anatomical landmarks for performing CSF collection Perform CSF collection on a standing, sedated horse Lab 11: Equine lateral thoracic vein catheterization <ul style="list-style-type: none"> Identify the lateral thoracic vein with ultrasound in standing horse Place over-the-wire catheter in the lateral thoracic vein in standing horse Place epidural catheter in standing sedated horse Lab 12: Ophthalmology Procedures <ul style="list-style-type: none"> Perform complete equine ophthalmic exam including tonometry, fluorescein staining, collection of cytologic samples, and ocular ultrasonography Perform auriculopalpebral and frontal nerve blocks Place dorsal and ventral sub-palpebral lavage systems Flush the nasolacrimal duct (time permitting) 	Bertin, Taylor, Jamieson, Trsan, Stapley, Townsend, Morgan
5:45 PM	Shuttle to hotel	

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DAY 4 Sunday, September 28		
Time (Eastern Time)	Topic	Presenter(s)
7:30 am	Shuttle to Brunner Equine Hospital (Purdue University)	
8:00-9:00 am	Principles of Technical Large Animal Emergency Rescue Training (TLAER) <ul style="list-style-type: none"> Understand and assess common TLAER situations encountered by practitioners and technicians (field and clinical scenarios) Manage safety and appropriate PPE, consider people positioning, extensions of the arm, physical restraint of recumbent animals, improve recovery times, make efficient decisions and work as a team Overview of manipulations, equipment and techniques utilized in common scenarios 	<i>Husted</i>
9:00-9:15 am	Break	
9:15-12:30 pm	<u>Labs – 50 Minutes per rotation</u> Lab 13: TLAER <ul style="list-style-type: none"> Perform manipulations (forward assist, backwards drag, sideways slide) of recumbent or trapped large animals, perform vertical lift and slinging from dorsal/lateral/posterior recumbency Lab 14: Perform Postmortem Procedures <ul style="list-style-type: none"> Perform AO CSF collection, intrathecal lidocaine euthanasia, tracheotomy, renal biopsy, thoracotomy, enucleation, etc. 	<i>Husted, Balser, all</i>
12:30-1:30 pm	Lunch and Course Wrap Up	

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